Qubit®
Fluorometer

probes.invitrogen.com/qubit
Welcome to the Qubit® Quantitation Platform—an efficient combination of sophisticated, accurate, and highly sensitive fluorescence-based quantitation assays with the most user-friendly fluorometer available. Because the Quant-iT™ assay kits use state-of-the-art dyes selective for dsDNA, RNA, and protein, you don’t need to worry about contaminants in your sample affecting the quantitation. And because the Qubit® fluorometer uses the very latest in illumination and detection technology for the highest sensitivity, you can use as little as 1 µL of sample and still get great results, even with very dilute samples. So plug in your new Qubit® fluorometer, grab your samples, and experience higher confidence in downstream applications.

To get the most out of your new Qubit® fluorometer, be sure to register online at the Qubit® website (probes.invitrogen.com/qubit). Registering your instrument ensures that you will receive notifications of software upgrades and information on new assays for use with the Qubit® fluorometer.
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Inspection and Setup

Upon receiving your Qubit® fluorometer, please inspect everything carefully and make sure all accessories are present. All shipments include:

- Qubit® fluorometer
- universal power cord with outlet adaptors, suitable for use with either a 110V or 220V electrical supply
- instruction manual
- abbreviated protocol card

1. Position the instrument on a flat, dry surface.
2. Plug one end of the power cord into the Qubit® fluorometer. To the other end attach the correct plug adaptor, based on the electrical outlet configuration in your country.
3. Plug the power cord into the electrical outlet. Be sure to use only the power cord provided with your instrument. Powering the instrument with an unapproved power supply may damage the instrument.
4. The instrument is automatically powered on when it is plugged in. To power down the Qubit® fluorometer, highlight the Power OFF command on the HOME screen and press GO. The Qubit® fluorometer has a sleep mode that is triggered after 60 minutes of inactivity. To recover from the sleep mode, press any of the four buttons on the instrument. Recovery from sleep mode returns the user to the same screen present when sleep mode commenced. After 10 hours of inactivity, the unit will power down automatically. To recover after powering down, press any of the four buttons on the instrument. Recovery from powered down mode returns the user to the Welcome screen.

Registering Your Qubit® Fluorometer

Visit probes.invitrogen.com/qubit to register your instrument. You will be asked to supply the serial number, your name, and your contact details. Registering your instrument ensures that you will receive notifications of software upgrades and information on new assays for use with the Qubit® fluorometer.
Installing the Qubit® Data Logger

The Qubit® fluorometer is designed for stand-alone use and does not require a connection with a PC. If data transfer to your PC is desired, you will need to install the Qubit® Data Logger. Visit the web address below and follow the instructions. When the software is installed, the Qubit® Data Logger icon will appear on your desktop. The requirements for download are:

**USB A/mini B 5-pin cable**
available from Invitrogen (Cat. no. Q32858) or from most office supply retailers

**PC System requirements**
- USB port (1.1 or later)
- Windows XP

**Location of downloadable software**
probes.invitrogen.com/qubit
Using the Qubit® Fluorometer

Critical Quant-iT™ Assay Considerations

Assay Temperature
The Quant-iT™ assays were designed to be performed at room temperature (22–28°C), as temperature fluctuations can influence the accuracy of the assay (Figure 1 on page 10). To minimize temperature fluctuations, store all kit reagents at room temperature and insert all assay tubes into the Qubit® fluorometer only for as much time as it takes for the instrument to measure the fluorescence, as the Qubit® fluorometer can raise the temperature of the assay solution significantly, even over a period of a few minutes. Do not hold the assay tubes in your hand before reading, as this will warm the solution and result in a low reading.

Incubation Time
To allow the Quant-iT™ assay to reach maximum fluorescence, incubate the tubes for the DNA and RNA assays for 2 minutes after mixing the sample or standard with the working solution. After this incubation period, the fluorescence signal is stable for 3 hours at room temperature, for all nucleic acid assays except the Quant-iT™ ssDNA assay, which is stable for up to 30 minutes. The Quant-iT™ protein assay requires 15 minutes’ incubation for a stable signal. For greatest accuracy of the protein assay, the incubation time of the samples should be within 10 minutes of the incubation time of the standards.

Photobleaching of the Quant-iT™ Reagent
The Quant-iT™ reagents exhibit high photostability in the Qubit® fluorometer, showing <0.3% drop in fluorescence after 9 readings and <2.5% drop in fluorescence after 40 readings. It is important to remember, however, that if the assay tube remains in the Qubit® fluorometer for multiple readings, a temporary reduction in fluorescence will be observed as the solution increases in temperature (see General Considerations for Analysis, Figure 1). (The temperature inside the Qubit® fluorometer may be as much as 3°C above room temperature after 1 hour.) For this reason, if you want to perform multiple readings of a single tube, remove the tube from the instrument and let it equilibrate to room temperature for 30 seconds before taking another reading.
Calibrating the Qubit® Fluorometer

For each assay, you have the choice to run a new calibration or to use the values from the previous calibration. As you first use the instrument, you should perform a new calibration each time. As you become familiar with the assays, the instrument, your pipetting accuracy, and significant temperature fluctuations within your laboratory, you should determine the level of comfort you have using the calibration data stored from the last time the instrument was calibrated. Remember also that the fluorescence signal in the tubes containing standards and the samples is stable for no longer than 3 hours. See Figure 2 in General Considerations for Analysis for an example of the calibration curve used to generate the quantitation results.

Assay Tubes for Use with the Qubit® Fluorometer

Only thin-wall, clear 0.5 mL PCR tubes are appropriate for use in the Qubit® fluorometer. Acceptable tubes include Qubit® assay tubes (Invitrogen Cat. no. Q32856, 500 tubes) or Axygen PCR-05-C tubes (VWR, part number 10011-830). The assay volume must be 200 µL for an accurate read.

Connecting the Qubit® Fluorometer to PC for Data Transfer (Optional)

It is not necessary to connect the Qubit® fluorometer to a PC. Quantitation results are displayed directly on the screen for you to record. If data transfer is desired, you need to have:

1. Qubit® Data Logger installed on your PC (see Installing the Qubit® Data Logger, above)
2. Qubit® fluorometer connected to the PC via the USB A/mini B 5-pin cable

The Qubit® fluorometer should be connected to the PC before the Qubit® fluorometer is powered on. When the Qubit® fluorometer is powered on, the PC may then let you know that it has recognized a peripheral device. As you proceed through the assay, the open spreadsheet will be updated with the data for standards and samples (data transfer is initiated when the first standard is read). At the end of the experiment, don’t forget to save the spreadsheet that contains the data for that session.
Selecting a Quant-iT™ Assay

1. If the fluorometer is powered down or in sleep mode, press any of the four buttons to power up the fluorometer.

   **Note:** If the instrument was in sleep mode, pressing any of the four buttons returns you to the last screen viewed before sleep mode commenced. To get to the HOME screen after reawakening, press HOME.

2. On the HOME screen, use the ▲ or ▼ key to highlight the assay of choice.

3. When the Quant-iT™ assay you would like to run is highlighted, press GO.

Calibrating the Qubit® fluorometer

For each assay, you have the choice to run a new calibration or to use the values from the previous calibration. See *Calibrating the Qubit® Fluorometer*, above. Highlight either **Run new calibration** or **Use last calibration** and press GO.

   **Note:** When using the fluorometer for the first time, there will not be an option to **Use last calibration**.

Running a New Calibration

Calibration of the Qubit® fluorometer requires the preparation of the appropriate standard solutions. For instructions on the preparation of these standards, see the instruction manual that accompanies the assay you are using.

   **Note:** It is important to prepare standards in the appropriate 0.5 mL assay tubes. Acceptable tubes include Qubit® assay tubes (500, Invitrogen Cat. no. Q32856) or Axygen PCR-05-C tubes (VWR, part number 10011-830). The assay volume must be 200 µL for an accurate read.

1. Insert Standard #1 and press GO. Ensure you use the Standard #1 appropriate for the assay you are performing.

2. The reading will take approximately 5 seconds.

3. Insert Standard #2, and press GO. Ensure you use the Standard #2 appropriate for the assay you are performing.

4. The Quant-iT™ protein assay requires a three-point calibration, so if you are performing this assay, a prompt to insert Standard #3 will appear on the screen. After Standard #3 is inserted, press GO.
5. The calibration is complete after Standard #2 is read (or after Standard #3 is read for the Quant-iT™ protein assay). If you get a “Standards incorrect” message, you have made an error. Checking the calibration values may help you to determine the source of your error. See Calibration Information, below.

**Using the Last Calibration**

You can choose to apply the previous calibration to your sample readings by pressing **GO** when **Use last calibration** is highlighted on the calibration selection screen. You are then directed to insert the assay tube containing your sample.

**Reading Samples**

It is important to prepare the sample correctly before a reading is taken. For instructions on the preparation of your samples, see the instruction manual that accompanies the assay you are using.

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**Note:** It is important to prepare samples in the appropriate 0.5 mL assay tubes. Acceptable tubes include Qubit® assay tubes (500, Invitrogen Cat. no. Q32856) or Axygen PCR-05-C tubes (VWR, part number 10011-830). The assay volume must be 200 µL for an accurate read.

1. After the calibration is complete or the values from the previous calibration have been accepted, insert a sample and press **GO**.

2. The measurement will take approximately 5 seconds. Upon the completion of the measurement, the result will be displayed on the screen. The number displayed is the concentration of the nucleic acid or protein in the assay tube.

3. Calculate the concentration of your original sample (see **Calculating the Concentration of Your Sample**, below) Alternatively, choose **Calculate sample concentration** to have the Qubit® fluorometer perform this calculation for you.

4. Record the reading.

5. Remove the sample from the instrument, insert the next sample, and press **GO**.

6. Repeat sample readings until all samples have been read.

7. If you did not choose **Calculate sample concentration**, calculate the concentration of your original sample by using the equation supplied below.
Calculating the Concentration of Your Sample

The Qubit® fluorometer gives values in either µg/mL or ng/mL. This value corresponds to the concentration after your sample was diluted into the assay tube. To calculate the concentration of your sample, use the following equation:

\[
\text{Concentration of your sample} = \text{QF value} \times \left( \frac{200}{x} \right)
\]

where:

- \( \text{QF value} \) = the value given by the Qubit® fluorometer
- \( x \) = the number of microliters of sample you added to the assay tube

This equation generates a result with the same units as the value given by the Qubit® fluorometer (i.e., if the Qubit® fluorometer gave a concentration in µg/mL, the result of the equation will be in µg/mL).

Calibration Information

The Check Calibration screen displays the raw values for Standard #1, Standard #2, Standard #3 (if applicable), and the last measurement taken for a given assay. These values can assist you in making a judgment regarding the performance of the Qubit® fluorometer and whether or not the samples you used were within range. For the Quant-iT™ dsDNA BR, Quant-iT™ dsDNA HS, Quant-iT™ ssDNA, and Quant-iT™ RNA assays, the reading given by Standard #2 should be much higher than that of Standard #1, and the reading of your sample should fall somewhere between that of the two Standards. For the Quant-iT™ protein assay, the reading given by Standard #3 should be much higher than that of Standard #2, and the reading of your sample should be no more than 50% greater than Standard #3.

1. To access the Check Calibration screen, press HOME from any screen.
2. Use the ▲ or ▼ key to highlight the Check Calibration option and press GO.
3. On the Check Calibration screen, use the ▲ or ▼ key to select the assay for which you want calibration information and press GO.
4. The raw values will be displayed on the screen.
Effect of Temperature on the Quant-iT™ Assays

The Quant-iT™ assays were designed to be performed at room temperature (22–28°C), as temperature fluctuations can influence the accuracy of the assay (Figure 1). To minimize temperature fluctuations, store all kit reagents at room temperature and insert all assay tubes into the Qubit® fluorometer only for as much time as it takes for the instrument to measure the fluorescence, as the Qubit® fluorometer can raise the temperature of the assay solution significantly, even over a period of a few minutes. Do not hold the assay tubes in your hand before reading, as this will warm the solution and result in a low reading.

Figure 1—Effect of temperature on the Quant-iT™ dsDNA BR assay. Quant-iT™ dsDNA HS, Quant-iT™ ssDNA, Quant-iT™ RNA, and Quant-iT™ protein assays show similar sensitivities over the same range.
How the Qubit® Fluorometer Calculates Concentration

The Qubit® fluorometer generates concentration data based on the relationship between the two Standards used in calibration (three for the Quant-iT™ protein assay). The plot below shows the line corresponding to the curve-fitting algorithm (a modified Hill plot) used in the calculation of concentration data for the Quant-iT™ RNA assay. For reference, the positions of the Standards and a set of data points from an actual experiment are shown superimposed onto the line. This plot demonstrates that the curve-fitting algorithm gives accurate values for quantitation.

Figure 2—The curve-fitting algorithm used to determine concentration in the Quant-iT™ RNA assay. Data for the Quant-iT™ dsDNA BR, Quant-iT™ dsDNA HS, Quant-iT™ ssDNA, and Quant-iT™ protein assays are generated by a similar algorithm.
Troubleshooting

Handling Samples
1. The calibration standards included in the Quant-iT™ RNA and Quant-iT™ RNA BR Assay Kits are high-quality rRNA standards. The integrity and concentration of these standards is critical to the optimal performance of the Quant-iT™ RNA assay. As such, we highly recommend treating the rRNA standards as you would any other precious RNA. Use appropriate RNAse-free handling techniques, including RNAse-free gloves, pipette tips, and tubes. Keep the tube lids closed whenever possible; do not touch the pipet to the inside wall of the tube when withdrawing a sample, and return the rRNA standard to the refrigerator as soon as possible after use.

2. Ensure that the assay tubes are at room temperature at the time the reading is taken. Do not hold assay tubes in your hand and do not leave assay tubes in the Qubit® fluorometer for longer than it takes to read the fluorescence. See Effect of Temperature on the Quant-iT™ Assays on page 10.

3. Be careful not to spill sample into the sample chamber. Promptly wipe any spills.

4. The Quant-iT™ assays are very sensitive and even small amounts of material from a previous sample may result in errors. Use a clean 0.5 mL PCR tube for each reading.

5. The tube MUST BE CLEAN AND DRY on the outside when taking readings. Moisture and condensation on the tube surface can lead to reading errors.

6. Minute bubbles in samples will cause errors in readings. Be sure not to introduce bubbles into samples. Slight tapping on the tube wall or brief centrifugation will often help dissipate bubbles.

Low Reading
1. Your sample is out of range. Use a sample that is more concentrated or use a lower dilution (i.e., 20 µL in 180 µL instead of 10 µL in 190 µL).

2. View the Check Calibration screen on the Qubit® fluorometer to confirm that the values for the samples fall between the values of the standards (see Calibration Information on page 9).

3. Ensure that you have prepared the Quant-iT™ working solution correctly (1:200 dilution using the buffer provided in the kit).
4. Ensure that you have prepared the standard tubes correctly (10 µL of each standard in 190 µL of Quant-iT™ working solution).

5. Ensure the tubes are filled to at least 200 µL (standards and samples).

6. Ensure that the reagents and standards you are using are not older than the expiration dates assigned by the manufacturer.

7. The Quant-iT™ stock solution should be protected from light as much as possible.

8. Select the correct Qubit® fluorometer assay for the Quant-iT™ assay you are performing and calibrate the Qubit® fluorometer correctly. Standards must be used in the correct order.

9. Ensure that the assay is operated entirely at room temperature.

**High Reading**

1. Your sample is out of range. Use a sample that is less concentrated.

2. View the Check Calibration screen on the Qubit® fluorometer to confirm that the values for the samples fall between the values of the standards.

3. Ensure that the lid is closed while reading standards and samples.

4. Prepare samples and standards according to the instructions in the Quant-iT™ assay kit you are using.

5. Ensure that the assay is operated entirely at room temperature.

**Standards Incorrect**

If you get a “Standards incorrect” message, you have made an error. Accessing the Check Calibration screen will allow you to view the raw values for Standard #1, Standard #2, Standard #3 (if applicable), and the last measurement taken for a given assay. These values can assist you in making a judgment regarding the performance of the Qubit® fluorometer and whether or not the samples you used were within range. See *Calibration Information* on page 9 for instructions for accessing the Check Calibration screen.
Warranty

Invitrogen warrants that this product will be free from defects in material and workmanship for a period of one (1) year from date of purchase. If a defect is present, Invitrogen will, at its option, repair, replace, or refund the purchase price of this product at no charge to you, provided it is returned during the warranty period. This warranty does not apply if the product has been damaged by accident, abuse, misuse, or misapplication, or from ordinary wear and tear. For your protection, items being returned must be insured against possible damage or loss. Invitrogen cannot be responsible for damage incurred during shipment of a repair instrument; It is recommend that you save the original packing material in which the instrument was shipped. This warranty shall be limited to the replacement of defective products. IT IS EXPRESSLY AGREED THAT THIS WARRANTY WILL BE IN LIEU OF ALL WARRANTIES OF FITNESS AND IN LIEU OF THE WARRANTY OF MERCHANTABILITY.
Obtaining Service

Contact Invitrogen Technical Service (in the USA and Canada call 1-800-955-6288 or send an email to probestech@invitrogen.com) and describe the problem(s) you are experiencing. Carry out any suggested modifications or tests. DO NOT ship an instrument to us without prior telephone or email contact. Often the problem is a relatively simple one that you can solve yourself with our direction.

If it is determined by the Invitrogen Technical Service representative that the instrument should be returned for repair, a Decontamination Certificate will be faxed to you. Once you have completed the certificate, signed it, and returned it to the Invitrogen Technical Service representative, a Return Authorization number will be assigned and you will receive instructions for the return.

If you are in the United States, and the instrument is under warranty, Invitrogen will repair or replace the unit, and pay for return shipment. If the instrument is not under warranty, Invitrogen will give you a cost estimate before repairing the unit. Repair and shipping costs both ways are your responsibility if the instrument is not under warranty.

If you are outside the United States and have purchased the instrument from an authorized Invitrogen distributor, contact that distributor directly. If you purchased directly from Invitrogen, you are welcome to send the instrument for free repair within the limits of the warranty, using the procedure described above. If it is not under warranty, we will repair it on a time-and-materials basis. We cannot, however, pay for shipping, duties, and documentation costs outside the continental United States.
### Product List

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<td>Q32860</td>
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<tr>
<td>Qubit® Quantitation Lab Starter Kit (includes 5 Qubit® fluorometers and 5 Quant-iT™ assay kits)</td>
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<tr>
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<tr>
<td>Quant-iT™ dsDNA BR Assay Kit, 100 assays <em>2-1000 ng</em> <em>for use with the Qubit® fluorometer</em></td>
<td>Q32850</td>
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<td>Q32853</td>
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<tr>
<td>Quant-iT™ dsDNA HS Assay Kit, 100 assays <em>0.2-100 ng</em> <em>for use with the Qubit® fluorometer</em></td>
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<td>Quant-iT™ ssDNA Assay Kit, 100 assays <em>1–200 ng</em> <em>for use with the Qubit® fluorometer</em></td>
<td>Q10212</td>
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<td>Quant-iT™ RNA Assay Kit, 100 assays <em>5-100 ng</em> <em>for use with the Qubit® fluorometer</em></td>
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<td>Quant-iT™ RNA BR Assay Kit, 100 assays <em>20–1000 ng</em> <em>for use with the Qubit® fluorometer</em></td>
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<th><strong>Qubit® Accessory Products</strong></th>
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<td>Qubit® assay tubes, 500 tubes</td>
<td>Q32856</td>
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<td>Qubit® USB cable</td>
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<td>Qubit® international power cord <em>replacement</em></td>
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# Instrument Specifications

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<tr>
<th>Specification</th>
<th>Description</th>
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<tr>
<td>Size</td>
<td>4.5” × 6.5” × 1.8” (11.4 cm × 16.6 cm × 4.6 cm)</td>
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<tr>
<td>Weight</td>
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<tr>
<td>Dynamic range</td>
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<td>Operating temperature</td>
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<tr>
<td>Light sources</td>
<td>Blue LED (max ~460 nm) Red LED (max ~635 nm)</td>
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<td>Excitation filters</td>
<td>Blue 400–490 nm Red 570–645 nm</td>
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<tr>
<td>Emission filters</td>
<td>Green 510–580 nm Red 655–725 nm</td>
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<td>Detectors</td>
<td>Photodiodes: measurement capability from 300–1000 nm</td>
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<td>Tube type</td>
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